# Phylogenetic Analysis of a Novel Molecular Isolate of Spotted Fever Group Rickettsiae from Northern Peru

# Candidatus Rickettsia andeanae

ABSTRACT: Phylogenetic analysis of five rickettsial genes (17-kDa gene, gltA, ompB, ompA, and sca4) from two molecular isolates of Candidatus Rickettsia andeanae from two ticks (Amblyomma maculatum and Ixodes boliviensis) collected from two domestic horses living in two separate locations in northern Peru (Coletas and Naranjo) was conducted to more clearly characterize this recently reported novel spotted fever group (SFG) rickettsia. Following nested polymerase chain reaction (PCR) amplification of the 17-kDa gene, gltA, ompB, ompA, and sca4, amplicons were purified, sequenced, and compared to those downloaded from GenBank. Phylogenetic analyses of the Candidatus Rickettsia andeanae sequences generated from 17-kDa gene (483 bp), gltA (1185 bp), ompA (1598 bp), ompB (4839 bp), and sca4 (2634 bp) demonstrated that they aligned strongly with those of SFG rickettsiae. Moreover, the sequences of these five genes most closely aligned with the following rickettsiae: ompA: Rickettsia sp RpA4 (98.03%), R. sp DnS28 (97.90%), and R. rhipicephali and R. massiliae (97.11%); ompB: R. aeschlimannii (97.22%), R. rhipicephali (97.20%), and R. sp Bar 29 (97.10%); and sca4: R. massiliae (97.8%), R. rhipicephali, and R. slovaca (97.7%). These results from the additional phylogenetic analyses of Candidatus Rickettsia andeanae confirm its inclusion within, and distance and uniqueness from, other known SFG rickettsiae.

KEYWORDS: Peru; ticks; spotted fever rickettsiae; phylogenetic analysis

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#### INTRODUCTION

Between May and October 2002, a number of febrile cases, including two deaths, were reported in an area around the town of Sapillica in northern Peru. A joint investigation with representatives of the Peruvian Ministry of Health and Naval Medical Research Center Detachment, Lima, documented a high prevalence of antibodies to Rickettsia and Leptospira species. During the investigation nucleic acid preparations extracted from blood samples obtained from febrile humans and peri-domestic rodents, and from fleas and ticks from individual domestic and wildtrapped animals, were determined by PCR as specific for four rickettsial genes, including: the Rickettsia genus-specific17-kDa outer membrane antigen gene (17kDa gene), the citrate synthase gene (gltA), and the outer membrane protein A (ompA) and B (ompB) genes to contain evidence for the presence of SFG rickettsiae.<sup>2</sup> Initially, all samples were tested for presence of the 17-kDa gene, and if detected determination of whether the amplicon was from a typhus or spotted fever group rickettsia was made. Prevalence as determined from these tests demonstrated that 24% of febrile humans (4/17), 8.3% of trapped rodents (2/24), 1.6% of flea pools (1/59), and 25% of ticks (4/16) were positive for SFG agents. Further analysis of the molecular isolates utilizing PCR specific for the gltA, ompA, and ompB genes and sequencing the subsequent amplicons determined the phylogenetic relationships of the molecular isolates. In addition to detecting R. felis in flea triturates, we identified a new SFG family member molecularly isolated from two tick samples (tick 124 and tick 163).<sup>2</sup> Due to the more than 3% difference in sequence identity between a 2,484 bp sequence segment of ompB from the novel agent and those SFG rickettsiae sequences listed in GenBank, it was proposed that the new agent be named Rickettsia andeanae. Herein, we describe further the characterization of Candidatus Rickettsia andeanae based upon phylogenetic analysis of five rickettsial genes. Four of the genes have been previously assessed (17-kDa gene 434 bp; gltA 381 bp; ompA 540 bp; and ompB 2,484 bp)<sup>2</sup> however, for this project larger amplicons from 17-kDa gene (483 bp), gltA (1185 bp) ompA (1,589 bp), and ompB (4,839 bp), and 2,634 bp of sca4 (120 kDa cytoplasmic protein gene; geneD) were analyzed and the results described in this report confirmed Candidatus Rickettsia andeanae inclusion within, and distance and uniqueness from other known SFG rickettsiae.

#### MATERIALS AND METHODS

Two molecular isolates of *Candidatus* Rickettsia andeanae from two ticks (*Amblyomma maculatum* and *Ixodes boliviensis*) collected from domestic horses living in two separate locations in Northern Peru (Coletas and Naranjo)<sup>2</sup> were evaluated by multilocus sequence typing to determine their relationship to other rickettsiae. Following amplification by standard polymerase chain reaction (PCR) of the 17-kDa gene and nested PCR amplification of *ompB*, *ompA*, and *sca4*, nucleic acid–derived from the two isolates, amplicons were purified and sequenced on an automated ABI Prism 3100 gene analyzer (Applied Biosystems, Foster City, CA) similar to that described previously.<sup>2</sup> However, new primers were used for PCR and sequencing in this study (Table 1). The sequences (both forward and reverse) were assembled with

TABLE 1. Primers for PCR, nested PCR, and sequencing

Target Gene	Primer	ner Sequence (5'-3')		
17-kDa antigen gene	Rp17kF1 <sup>ac</sup>	AATGAGTTTTATACTTTACAAAAT- TCTAAAAAACCA	532	
	Rr1175F <sup>ac</sup>	GCTCTTGCAACTTCTATGTT	434	
	Rr2608Rac	CATTGTTCGTCAGGTTGGCG		
gltA	CS1dF <sup>abc</sup>	ATGACTAATGGCAATAATAA	1254	
	CS1273R <sup>a</sup>	CATAACCAGTGTAAAGCTG		
	$CS1234R^{bc}$	TCTAGGTCTGCTGATTTTTTGTTCA		
	Rp CS877F <sup>bc</sup>	GGG GGC CTG CTC ACG GCG G	382	
	Rp CS1258R <sup>bc</sup>	ATTGCAAAAAGTACAGTGAACA		
ompA	190-3588F <sup>a</sup>	AACAGTGAATGTAGGAGCAG	1630	
	190-5238R <sup>abc</sup>	ACTATTAAAGGCTAGGCTATT		
	$RompA1F^{bc}$	GAATAACATTACAVGCYGGAGGAAG		
	$RrA657R^c$	TATTTGCATCAATCSYATAAGWA		
	RhoA4336F <sup>c</sup>	AGTTCAGGAAACGACCGTA		
ompB	RompB11F <sup>a</sup>	ACCATAGTAGCMAGTTTTGCAG	4887	
	RompB4887R <sup>ab</sup>	AGAGTACCTTGATGTGCRGTATAYT		
	$RompB2409F^{bc}$	CCGTAACATTAAACAAACAAGCTG	2479	
	RompB2553R <sup>bc</sup>	GAATTTTCAAAAGCAATYGTATCAGT	2553	
	$RAK1009F^{bc}$	ACATKGTTATACARAGTGYTAATGC		
	$RAK1452R^{bc}$	SGTTAACTTKACCGYTTATAACTGT		
	$120-2788F^{c}$	AAACAATAATCAAGGTACTGT		
	RompB3521F <sup>c</sup>	GATAATGCCAATGCAAATTTCAG		
	RompB4224F <sup>c</sup>	ACCAAGATTATAAGAAAGGTGATAA		
	$120-4346R^{c}$	CGAAGAAGTAACGCTGACTT		
	RompB3637R <sup>c</sup>	GAAACGATTACTTCCGGTTACA		
	$RompB3008R^c$	AATATCGCTGACGGTCAAGGT		
	$120-607F^{c}$	AATATCGCTGACGGTCAAGGT		
	$120-807R^{c}$	CCTTTTAGATTACCGCCTAA		
	$RR1595F^c$	GCCGGAGTTGTCCAATTATCA		
	RompB1902R <sup>c</sup>	CCGTCATTTCCAATAACTAACTC		
	RompB2012F <sup>c</sup>	GCCGGTACAAATTTAGGTAGTG		
sca4	D1f <sup>abc</sup>	ATGAGTAAAGACGGTAACCT	3069	
	D3069rabc	TCAGCGTTGTGGAGGGGAAG		
	RrD749F <sup>abc</sup>	TGGTAGCATTAAAAGCTGATGG		
	RrD2685Rabc	TTCAGTAGAAGATTTAGTACCAAAT	2706	
	RrD928Fbc	ATTTATACACTTGCGGTAACAC		
	RrD1826Rbc	TCTAAATKCTGCTGMATCAAT		
	RrD1713F <sup>c</sup>	CTCTGAATTAAGCAATGCGGAAA		
	T5DR1 <sup>c</sup>	CTGATAAAGCTGTAGCTGCATTA		
	D2338f <sup>c</sup>	GATGCAGCGAGTGAGGCAGC		
	D928r <sup>c</sup>	AAGCTATTGCGTCATCTCCG		

 $<sup>^</sup>a$ Primers used for PCR amplification;  $^b$ Primers used for nested PCR amplification;  $^c$ Primers used for sequencing.

		G	Length			Fournier and
Gene	Tick Sample	Sequence Length (bp)	to R. conorii	Position to ORF	Results (percent identity with closest neighbors)	colleagues Cutoff Values <sup>a</sup>
17-kDa	Tick124 Tick163	422 483	480	43~464 M30~464	98.49% to R. rickettsii; 98.28% to R. sibirica and R. conorii; 98.06% to R. rhipicephali; 97.84% to R. parkeri	b
gltA	Tick124 Tick163	354 1185	1308	810~1163 45~1229	99.42% to Rickettsia sp. DnS14; 99.34% to R.sp. RpA4, R. sibirica and R.sp. MC16, R.sp. BJ-90; 99.26% to R. parkeri, R. slovaca 99.17% to R. conorii, 99.12% to R. aeschlimannii; 99.09% to R. rhipicephali	99.9%
ompA	Tick124 and Tick163	1598	6066	2880~4464	98.03% to R.sp. RpA4; 97.90% to R.sp. DnS28; 97.84% to R. sp. DnS14; 97.11% to R. rhipicephali and R. massiliae	98.8%
ompB	Tick124 and Tick163	4839	4968	89~4927	97.22% to R. aeschlimannii; 97.20% to R. rhipicephali; 97.10% to R. sp. Bar 29; 97.08% to R. massiliae; 96.10% to R. slovaca	99.2%
sca4	Tick124 and Tick163	2634	3081	39~2672	97.8% to R. massiliae; 97.7% to R. rhipicephali and R. slovaca; 97.6% to R. aeschlimanii; 97.5% to R. conorii	99.3%

TABLE 2. Candidatus Rickettsia andeanae sequence identity with closest neighbors

Sequencher 4.0 software (Gene Codes Corp, Ann Arbor, MI) and compared to those downloaded from GenBank. Phylogenetic analyses (Neighbor Joining Best Tree and Bootstrap methods) were performed with MacVector 7.0 software (Accelrys, Inc, San Diego, CA).

#### RESULTS

## 17-kDa Gene and gltA

Comparing sequences of the *Candidatus* Rickettsia andeanae 17-kDa gene and *gltA* fragments with GenBank sequences indicate that *Candidatus* Rickettsia andeanae belongs within the genus *Rickettsia* by showing that the 483 bp 17-kDa gene segment had >97.5% identity with five SFG rickettsiae, but with no more then 98.5% identity with any rickettsial agent (Table 2). Similarly, the 1,185 bp *gltA* sequence fragment of *Candidatus* Rickettsia andeanne had 99.00–99.42% identity to ten SFG

rickettsiae, but no identity with any rickettsial agent >99.42% (TABLE 2). Thus, the sequencing data from the 17-kDa gene and *gltA* segments indicate that *Candidatus* Rickettsia andeanae is a rickettsial agent but not identical with any agent currently listed within GenBank.

#### ompA

Due to presence of *ompA* among SFG rickettsiae, and the lack of a complete *ompA* among TG rickettsiae, *R. bellii* and *R. canadensis*, the production of a PCR amplicon utilizing *ompA* specific primers corroborates the *gltA* data that suggests that *Candidatus* Rickettsia andeanae is closely related to SFG rickettsiae. In addition, phylogenetic analysis of a 1598 bp segment of *ompA* indicates that *Candidatus* Rickettsia andeanae has 98.03% identity with its closest neighbor, *Rickettsia* sp. RpA4, within the SFG (TABLE 2).

### ompB

Initial data from the phylogenetic analysis of *ompB* sequence (2,484 bp) from *Candidatus* Rickettsia andeanae showed that it was ≥3% divergent from other SFG rickettsiae.<sup>2</sup> In this investigation an *ompB* fragment almost twice as large (4,839 bp) was sequenced and evaluated for its relationship to other rickettsial (and non-rickettsial) sequences. It was determined that the larger segment also showed an approximately 3% divergence from the *ompB* sequences of the closest relations, which were determined to be SFG rickettsiae (TABLE 2). The most closely related rickettsiae based upon comparison of the 4,839 bp fragment of *ompB* were *R. aeschlimannii* (97.22% identity), *R. rhipicephali* (97.20% identity), and *R.* sp. Bar 29 (97.10% identity). The *Candidatus* Rickettsia andeanae *ompB* sequence showed only 87.7 and 87.3% identity with *R. prowazekii* and *R. typhi*, respectively.

#### sca4

Sequence comparisons of part or all of *sca4* have recently been described as a tool for rickettsial phylogenetic analysis because the presence of the gene among the various species of *Rickettsia* and due to its moderate sequence variability among the *Rickettsia*. Blast search with the 2,634 bp *sca4* segment sequence from *Candidatus* Rickettsia andeanae, showed that it was most closely related to SFG rickettsiae *R. massiliae* (97.8%), *R. rhipicephali* and *R. slovaca* (97.7%). Similar to the sequence comparison data discerned with *ompA* and *ompB*, *Candidatus* Rickettsia andeanae *sca4* sequence shows a close relationship to SFG rickettsiae, but is divergent enough to be considered a unique rickettsial species (TABLE 2).

#### DISCUSSION

Multilocus sequence typing (MLST) for the characterization of pathogenic and non-pathogenic microorganisms utilizes the sequences from several gene segments to characterize the genetic makeup of an isolate.<sup>4</sup> The sequences are compared to those already obtained for similar and dissimilar organisms. From the comparison a determination is made as to the relationship between the new isolate and known iso-

lates. This is very helpful in ascertaining the identity of an isolate(s) during an outbreak investigation, historical classification, or taxonomic relationships and has been used to characterize rickettsial agents.<sup>3,5</sup> In this investigation PCR amplified segments from five well known genes were used to validate the divergence of the novel isolate, *Candidatus* Rickettsia andeanae, from those rickettsiae previously characterized utilizing the criteria proposed by Fournier and colleagues.<sup>3</sup> The results presented herein confirm our earlier report<sup>2</sup> by extending the sequences assessed for four genes (17 kDa gene, *gltA*, *ompA*, ompB) and newly assessing the sequence of *sca4*. Collectively these results show the uniqueness of *Candidatus* Rickettsia andeanae, a novel agent that has been molecularly isolated from two different ticks collected from two different horses from two locations in northern Peru. However, an isolate has not been cultured in the laboratory and therefore the pathogenicity of this agent has not been ascertained. Collaborators in Peru are actively pursuing a course for cultivating this agent in their laboratory.

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### REFERENCES

- 1. BLAIR, P.J., G.B. SCHOELER, C. MORON, et al. 2004. Evidence of rickettsial and leptospira infections in Andean Northern Peru. Am. J. Trop. Med. Hyg. 70: 357–363.
- BLAIR, P.J., J. JIANG, G.B. SCHOELER, et al. 2004. Characterization of spotted fever group rickettsiae in flea and tick specimens from northern Peru. J. Clin. Microbiol. 42: 4961–4967.
- 3. FOURNIER, P-E., J.S. DUMLER, G. GILBERT, *et al.* 2003. Gene sequence-based criteria for identification of new *Rickettsia* isolates and description of *Rickettsia heilongjiangensis* sp. nov. J. Clin. Microbiol. **41:** 5456–5465.
- MAIDEN, M.C.J., J.A BYGRAVES, E. FEIL, et al. 1998. Multilocus sequence typing: a portable approach to the identification of clones within populations of pathogenic microorganisms. Proc. Natl. Acad. Sci. USA 95: 3140–3145.
- ISHIKURA, M., S. ANDO, Y. SHINAGAWA, et al. 2003. Phylogenetic analysis of spotted fever group rickettsiae based on gltA, 17-kDa, and rOmpA genes amplified by nested PCR from ticks in Japan. Microbiol. Immunol. 47: 823–832.